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Nucleosides, Nucleotides and Nucleic Acids

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Efficient Preparation of Cyclic 3',5'-Phosphoramidites and -Amidates of Antiviral and Antitumor 5-X-2'-Deoxyuridines (X = H, CH₃, I, F, CF₃, trans-CH=CHBr)

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To cite this Article Bentrude, Wesley G. , Khan, Mamunur R. , Saadein, M. R. and Sopchik, Alan E.(1989) 'Efficient Preparation of Cyclic 3',5'-Phosphoramidites and -Amidates of Antiviral and Antitumor 5-X-2'-Deoxyuridines (X = H, CH₃, I, F, CF₃, trans-CH=CHBr)', *Nucleosides, Nucleotides and Nucleic Acids*, 8: 7, 1359 — 1367

To link to this Article: DOI: 10.1080/07328318908054340

URL: <http://dx.doi.org/10.1080/07328318908054340>

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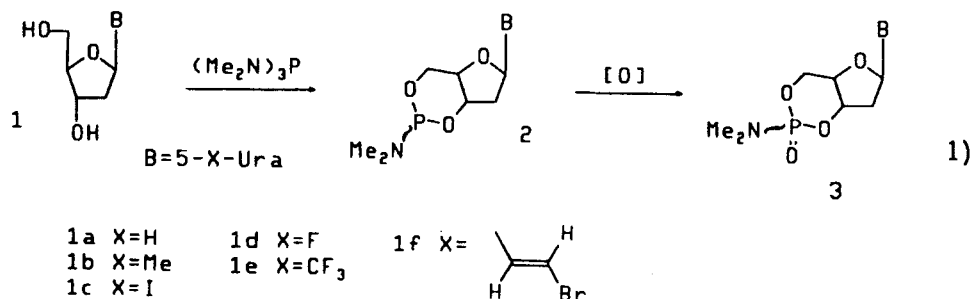
**EFFICIENT PREPARATION OF CYCLIC 3',5'-PHOSPHORAMIDITES AND
-AMIDATES OF ANTIVIRAL AND ANTITUMOR 5-X-2'-DEOXYURIDINES
(X = H, CH₃, I, F, CF₃, trans-CH=CHBr).**

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Abstract: Direct cyclization of the title nucleosides with (Me₂N)₃P followed by oxidation with N₂O₄ or *t*-BuOOH affords the individual cyclic 3',5'-phosphoramidate diastereomers shown to be isolable in 45-77% yields.

Neutral derivatives of nucleoside cyclic 3',5'-monophosphates have proven to be valuable in studies of enzyme active site mapping,² as mimics or antagonists of cAMP or cGMP,³ as possible antitumor or antiviral prodrugs,⁴ and in investigations of the energetics of the chair-twist equilibrium available to the six-membered phosphate ring.⁵ Nucleoside cyclic phosphoramidates (P(O)NR₂) have been of considerable interest in this regard and in addition, as the anilidates (P(O)NHPh), are very useful precursors of phosphorothioates and cyclic phosphate diesters which are chiral by virtue of isotopic oxygen substitution on phosphorus.⁶ A number of preparations of nucleoside cyclic 3',5'-phosphoramidates have been reported.^{6,7}

The potentially most straightforward access to the cyclic phosphoramidates of 2'-deoxyribonucleosides is by the way of reaction 1:

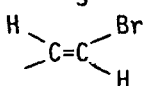


The phosphoramidite, 2, thus formed is an extremely valuable intermediate as has been demonstrated for thymidine (Scheme). Thus, phosphoramidite 2b has been shown previously to be converted nearly quantitatively to the corresponding phenyl^{5a} or methyl phosphite (4)⁸ from which cyclic phosphate triesters (5⁸ and 10^{5a}), methylphosphonates (6),⁹ phosphorothioates (7),¹⁰ and ¹⁷O- and ¹⁸O-labeled cyclic diesters (8)¹¹ can be readily prepared in high yields. Furthermore, 4 is obtained as a mixture of diastereomers which results in the formation of useful amounts of both diastereomers of products 5-9.

Unfortunately, up to now the yields of phosphoramidite 2b have never been reliably above 10-20%^{5a,8,12} which has precluded the application of these methods to more costly 2'-deoxyribonucleosides. We report here the preparation of the cyclic phosphoramidites of a series of 5-substituted 2'-deoxyribonucleosides (1a-1f) in good yields from 0.5-1.0 g amounts of nucleoside as evidenced by the ³¹P NMR spectra of the crude phosphoramidites (2). After oxidation, HPLC separation applied to a portion of the crude product gave the corresponding phosphoramidates (3) in 45-77% yields based on starting nucleoside. Useful amounts of the *cis* (relationship of Me₂N and base) as well as the predominant *trans* diastereomer were formed (See Table 1). It should be emphasized, however, that the *trans* phosphoramidite can be converted to the methyl phosphite (e.g. 4) with either the *cis* or *trans* isomer in excess and thus can yield *cis* phosphoramidate predominantly (4+9).^{7h,i} Thus, formation of *cis*-2 is only important when the *cis* phosphoramidate is required to be prepared directly (1 → 2 → 3). Nucleosides 1c-1f are all in clinical use or in various stages of testing as antivirals or antitumor agents.¹³ Reaction 1 and those of the Scheme provide an excellent approach, therefore, to potential prodrug forms of 1c-1f.¹⁴

The crucial aspects of the experimental procedure that ensure high yields are the use of carefully purified acetonitrile as solvent, thorough deoxygenation of the reaction, slow increase of the temperature of the reaction, and its control below 65 °C. The cyclization with 2'-deoxyuridine is sluggish unless tetrazole is added to catalyze the process. A greater percentage of *cis* phosphoramidate for direct oxidation to *cis*-3 can usually be obtained, at a small sacrifice in overall yield, by reduction of the reaction time. This approach gives minor amounts of what appear by ³¹P NMR to be uncyclized 3'- and 5'-

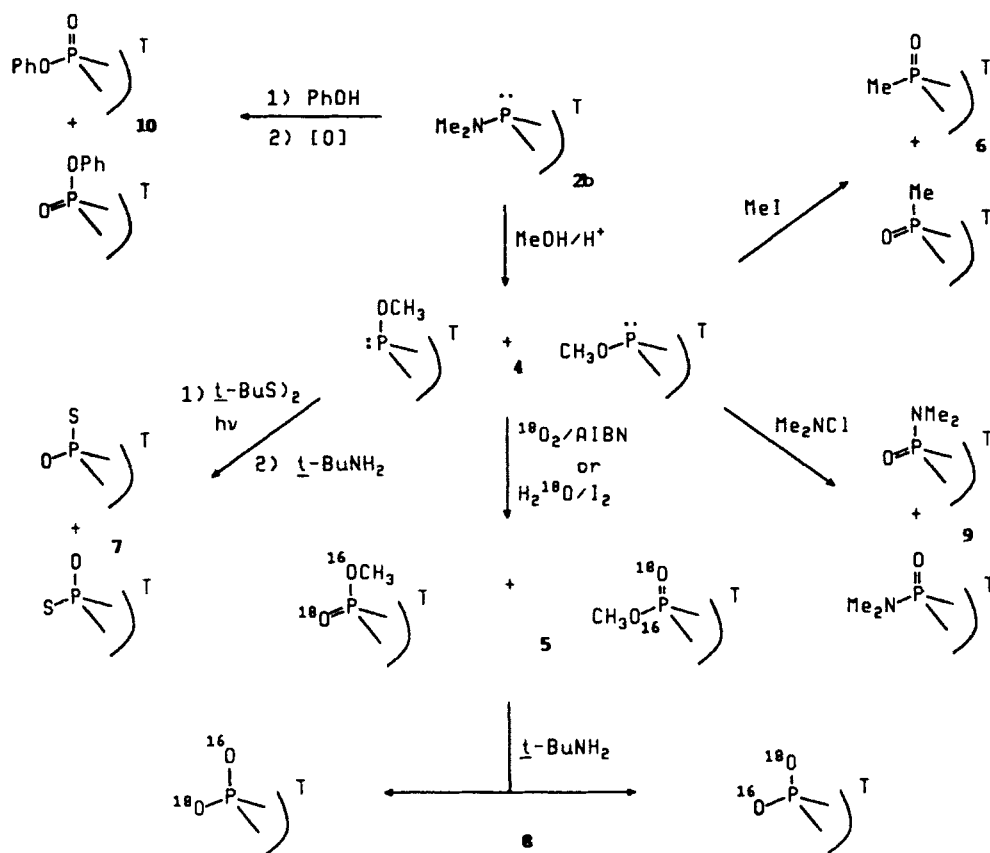
TABLE 1. Product Data for 5-X-2'-Deoxyuridine Cyclic 3',5'-Phosphoramidates.

Compd.	X	Total Isolated ^a Yield (%) of 3	$\delta^{31\text{P}}$ ^b		Ratio of ^c trans/cis
			cis	trans	
3a	H	45	7.69	9.43	83/17
3b	CH ₃	77	7.92	9.15	77/23
3c	I	64	7.80	9.26	78/22
3d	F	70	7.68	9.23	84/16
3e	CF ₃	55	7.73	9.27	86/14
3f		66	7.87	9.35	81/19

^aBy HPLC, 50-100 mg scale, Dynamax Macro SiO₂ column, single pass. Solvent system CH₂Cl₂/MeOH = 97/3 for compounds 3a-3c and CH₂Cl₂/MeOH = 98/2 for compounds 3d-3f. ^bAcetone-d₆ 121.3 MHz. Chemical shifts (ppm) are reported relative to external 85% H₃PO₄. ^cBy ³¹P quantitative NMR and/or HPLC.

phosphorodiamidites. These assumed precursors of 3 do not interfere, however, with workup of the reaction.

The structures of the individual diastereomers of 3a and 3c-3f were verified by their ¹H, ³¹P, and ¹³C NMR parameters on comparison to those of the known phosphoramidate, 3b.^{7f,9,15} The structures of both the cis and trans diastereomers of 3b have been proven by X-ray crystallography.^{7i,16} Cis and trans geometries were assigned to the individual diastereomers of 3a and 3c-3f on the basis of their relative ³¹P chemical shifts, as is well preceded.^{7f} Quantitative elemental analyses are given in Table 2. ¹H chemical shifts appear in Table 3. J_{HH} and J_{HP} values for the phosphate rings of 3a and 3c-3f are similar to the published values^{7h,5d} for 3b and will appear elsewhere in connection with conformational analysis of the phosphoramidate rings. Trends in



SCHEME

TABLE 2. Analytical Data for 3a and 3c-3f.^a

Compound	Formula		%C	%H	%P
3a	C ₁₁ H ₁₆ N ₃ O ₆ P	Calcd.	41.64	5.04	9.78
		Found	41.96	5.01	9.97
3c	C ₁₁ H ₁₅ N ₃ O ₆ PI	Calcd.	29.79	3.39	6.99
		Found	29.69	3.47	7.17
3d	C ₁₁ H ₁₅ N ₃ O ₆ PF	Calcd.	39.40	4.48	9.25
		Found	39.26	4.56	8.93
3e	C ₁₂ H ₁₅ N ₃ O ₆ PF ₃	Calcd.	37.40	3.89	8.05
		Found	37.30	4.06	7.89
3f	C ₁₃ H ₁₇ N ₃ O ₆ PBr	Calcd.	36.97	4.03	7.35
		Found	36.57	4.07	7.55

^aCarried out on 40/60 to 60/40 cis/trans mixtures.

TABLE 3. ^1H Chemical Shifts for the Diastereomers of 3a-3f in Acetone- d_6 at 26°. ^a

Compound	H1'	H2' ^b	H3'	H4'	H5'a	H5'b	H5	H6	NH	NMe ₂	Other
<u>cis</u> -3a	6.33	2.50-2.59	4.85	4.16	4.32	4.58	6.03	7.83	9.10	2.65	
<u>trans</u> -3a	6.35	2.52-2.64	4.86	3.93	4.46	4.50	5.63	7.74	7.99	2.67	
<u>cis</u> -3b ^d	6.40	2.64-2.67	4.94	4.20	4.31	4.61		7.51	c	2.68	1.84 ^e
<u>trans</u> -3b ^d	6.42	2.53-2.60	4.92	3.93	4.48	4.49		7.62	c	2.70	1.84 ^e
<u>cis</u> -3c	6.35	2.56-2.70	4.94	4.17	4.34	4.58		8.02	c	2.56	
<u>trans</u> -3c	6.33	2.58-2.75	4.94	3.94	4.47	4.52		8.12	c	2.67	
<u>cis</u> -3d	6.39	2.59-2.68	4.91	4.17	4.39	4.58		7.83	c	2.56	
<u>trans</u> -3d	6.35	2.53-2.65	4.98	3.94	4.48	4.54		7.97	10.47	2.67	
<u>cis</u> -3e	6.32	2.60-2.72	4.92	4.22	4.35	4.60		8.11	c	2.64	
<u>trans</u> -3e	6.30	2.58-2.74	4.92	3.99	4.50	4.54		8.16	c	2.66	
<u>cis</u> -3f	6.41	2.58-2.68	4.91	4.21	4.38	4.59		7.79	10.34	2.65	7.39, 6.81 ^f
<u>trans</u> -3f	6.42	2.56-2.65	4.89	3.96	4.47	4.53		7.81	c	2.67	7.41, 6.92 ^f

^aIn ppm downfield from internal TMS. ^b2H multiplet. ^cPeak not observed. ^dAt 300 MHz. ^e5-Me.^f5-CH=CHBr.

TABLE 4. ^{13}C NMR Parameters for 3a-3f in Acetone- d_6 at 75.5 MHz, 26°.

Compound	Chemical Shift (J_{CP} , Hz ^a)									
	C1'	C2' ^b	C3'	C4'	C5'	C2	C4	C5	C6	Me ₂ N
cis-3a ^e	87.4	36.7 (7.5)	78.4 (5.2)	74.6 (10.6)	69.4 (8.3)	151.0	164.1	102.8	143.4	36.2 (4.7)
trans-3a ^b	87.3	36.1 (8.5)	77.8 (4.2)	75.4 (4.7)	70.2 (7.7)	151.7	166.0	103.4	143.2	36.4 (5.0)
cis-3b	85.7	35.1 (7.6)	77.9 (5.2)	73.7 (11.3)	68.6 (8.4)	150.5	163.7	110.9	137.0	35.8 (3.7)
trans-3b	85.0	35.7 (8.4)	76.9 (4.0)	74.9 (4.5)	69.0 (7.6)	151.1	164.2	111.8	137.2	36.2 (4.6)
cis-3c	86.7	35.9 (7.1)	78.1 (5.2)	74.5 (11.1)	69.0 (8.4)	150.3	160.3	69.6	146.1	36.3 (3.6)
trans-3c	86.2	36.0 (8.4)	76.7 (3.7)	75.2 (4.1)	69.1 (7.6)	150.8	160.7	69.6	146.3	36.4 (4.7)
cis-3d ^e	86.1	35.9 (7.1)	78.0 (5.1)	74.5 (10.9)	69.0 (8.4)	149.3	157.3	141.3 (232.2) ^c	125.6 (34.2) ^d	36.3 (3.5)
trans-3d ^b	87.2	36.0 (8.5)	77.6 (4.2)	75.4 (4.6)	70.1 (7.8)	150.4	159.2	142.0 (232.4) ^c	126.8 (34.4) ^d	36.4 (5.1)
cis-3e ^e	88.1	36.6 (7.3)	78.0 (5.2)	74.8 (11.2)	69.0 (8.4)	149.7	158.9	143.4 (32.6) ^c	104.8 (5.8) ^d	36.3 (3.4)
trans-3e	87.5	36.0 (8.5)	76.5 (4.0)	75.4 (4.3)	68.9 (7.4)	150.1	159.2	143.4 (32.6) ^c	105.0 (5.9) ^d	36.2 (4.8)
cis-3f ^e	86.3	36.1 (7.1)	78.1 (5.2)	74.6 (11.3)	68.9 (8.2)	149.4	161.9	108.3	140.3	36.3 (3.7)
trans-3f	85.4	36.0 (8.2)	76.6 (3.9)	75.1 (4.3)	69.0 (7.5)	149.9	162.3	108.6	140.3	36.3 (4.5)

^a Acquisition times 2.1 s. Errors in J values ± 0.5 Hz.^b In CD₃OD.^c J_{C5F} .^d J_{C6F} .^e At 125.8 MHz.^f 5-Me.^g 5-CF₃.^h 5-CH=CHBr.

relative ^1H chemical shifts potentially useful in assigning cis and trans structures are evident in Table 2. ^{13}C parameters are recorded in Table 4. The relative ^{13}C chemical shifts for C3' and C4' correlate well with the cis or trans identity of the particular diastereomer as do J_{PC} values for C2'-C5'. These correlations were pointed out previously.^{7f}

EXPERIMENTAL

^1H NMR spectra were taken on a Varian VXR-500 spectrometer at 500 MHz. ^{13}C NMR spectra were obtained on a Varian XL-300 MHz spectrometer at 75.5 MHz and on a VXR-500 MHz at 125.8 MHz. ^{31}P NMR spectra were recorded on a Varian XL-300 instrument at 121.3 MHz.

Anhydrous solvents were obtained as follows: acetonitrile by successive distillations from calcium hydride, phosphorus pentoxide and then calcium hydride; dichloromethane by distillation first from phosphorus pentoxide and then from calcium hydride. Hexamethylphosphorus triamide was obtained from Aldrich Chemical Co. and distilled before use. Tetrazole from Aldrich Chemical Co. was sublimed prior to use. HPLC grade solvents were Omnisolve materials from EM Industries. A toluene solution of *t*-BuOOH from Fluka Chemie AG was used as received.

Preparation of 2b-2f and 3b-3f. Reactions were run on 0.5-1.0 g of nucleoside. The procedure for thymidine is typical. Thymidine (1.0 g, 0.41 mmol) was mixed with 100 mL of carefully purified acetonitrile in a flask equipped with a rubber septum. Argon was bubbled through the mixture for 15 min, and freshly distilled $(\text{Me}_2\text{N})_3\text{P}$ (0.67 g, 0.74 mL, 0.41 mmol), 99.9% pure by GLC, was added. The cloudy mixture was purged with argon for an additional 10 min. The flask was fitted with an argon-flushed condenser topped with a gas bubbler through which a slow flow of argon was maintained. The temperature of the reaction was slowly increased over a 1-2 h period and maintained at 63-65 °C for 18 h, during which time the reaction mixture became clear. Solvent was removed under oil pump vacuum to leave a white, foamy solid (1.25 g, 95% crude yield). ^{31}P NMR (CDCl_3) showed peaks at δ 146.1 (trans) and 140.1 (cis), ratio 75/25 (trans/cis), which comprised at least 90% of the total peak area. The crude product was dissolved in 50 mL of purified CH_2Cl_2 and maintained under argon while 1 mL of *t*-BuOOH (approximately 3 M solution in toluene, diluted with 5 mL of CH_2Cl_2) was added dropwise at 0 °C with stirring. Solvent evaporation left a foamy, white product (1.15 g,

0.34 mmol, 83% crude yield based on thymidine). The crude phosphoramidate (100 mg) was purified by HPLC in one pass on a Dynamax Macro SiO₂ column (25 cm x 21.4 mm; CH₂Cl₂/MeOH, 97/3, flow rate 10 mL/min) to give 71 mg of trans-3b and 20 mg of cis-3b.¹⁷ (Total yield based on 100 mg of crude product, 77%). As an alternative oxidation, the cyclic phosphoramidite was dissolved in 30 mL of CH₂Cl₂ and oxidized at -70° C with a saturated CH₂Cl₂ solution of N₂O₄ added dropwise until a pale yellow-green color persisted. N₂O₄ was the reagent of choice except for X = Me or I for which t-BuOOH gave equally good yields.

Preparation of 2a and 3a. These products were obtained by a procedure identical to the above except that 20 mg of tetrazole was added to a 500 mg-scale reaction.

Acknowledgment. Support of this research by Grant CA 11045 from the N.C.I. of the P.H.S. is gratefully acknowledged. For starting nucleosides we thank Dr. W.E. Scott, Hoffmann LaRoche Co. (5-F-dUrd), Dr. H.J. Schaeffer, Burroughs Wellcome Co. (5-CF₃-dUrd), and Dr. E. De Clercq (BVDU).

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Received January 22, 1989.